

# ASSESSING TEMPORAL SHIFTS IN SOIL MICROBIAL DIVERSITY IN PESTICIDE- EXPOSED AGRICULTURAL ECOSYSTEMS

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## Abstract

Pesticides may change soil physical, chemical, and microbiological activity (enzyme activities, microbial populations). Limited literature on enzyme-based soil quality monitoring during pesticide application. Existing literature on pesticide effects on soil enzyme activity does not investigate all enzymes. The study examined how carbofuran and paraquat affected soil biochemistry in India. This study utilized traditional strategies to survey the effect of pesticides on soil natural carbon, catalyst movement, and microbial populaces when applied at approved focuses. In particular, the chemicals being scrutinized included amylase, invertase, protease, urease, phosphatase, and dehydrogenase. Complete heterotrophic microbes, organisms, actinomycetes, nitrifying microorganisms, and phosphate solubilizers were measured through spread plate counting. The consequences of this exploration uncovered contrasts in compound action between soil treated with carbofuran and paraquat. Dehydrogenase action was viewed as upgraded by the treatment, while urease action displayed lower levels contrasted with different chemicals. The degrees of soil natural carbon showed variety over the span of the review. Quite, the microbial populaces bit by bit expanded because of their capacity to process and use pesticides as a wellspring of carbon and energy for a brief time. Fundamentally, a p-worth of 0.05 showed eminent qualifications in soil natural carbon, compound action, and microbial numbers. In

rundown, this study exhibited that the sort of pesticide, when applied at endorsed field application rates, transiently affects microbial populaces and compound action. Changes in soil microbe populations, activity, and variety may indicate soil fertility and quality.

**Keywords:** *Agricultural, Microbial Diversity, Pesticides, Ecosystems, Soil*

## 1. INTRODUCTION

The global population is currently facing a significant challenge in terms of population growth, necessitating an increased production of food to meet the growing demand. With approximately 75% of the Earth's surface being covered by water, there is a significant constraint on the availability of land for the expansion of food production. Hence, the sole means of augmenting agricultural output within the confines of limited land resources is by means of intensive agriculture, which represents the exclusive avenue for enhancing crop yield. The judicious application of inorganic fertilizers and pesticides does not pose a significant threat to the natural environment. However, the escalating demand for these compounds has resulted in their excessive and indiscriminate utilization, driven by the objective of achieving rapid and heightened output. This unrestricted usage is anticipated to yield enduring consequences. Based on available information, it has been observed that the environment's general well-being and the soil's health are experiencing adverse effects due to the indiscriminate use of these chemicals. Pesticides are a category of compounds employed for the purpose of pest management in both soil and plants. While their primary function is to safeguard crops from pests, it is worth noting that their usage might potentially have adverse effects on soil health.

The alterations arising from disturbances induced by the over utilization of agrochemicals, encompassing a diverse range of pesticides and inorganic fertilizers. The use of pesticides have the capacity to induce modifications in the microbial communities residing within the soil, hence impacting several crucial processes responsible for maintaining soil health. Enzymatic movement is a basic part of the regular habitat, assuming an essential part in starting compound responses. The presence of enzymes in the soil is closely associated with the microbial population. Consequently, the use of pesticides might potentially have detrimental impacts on biological processes and enzymatic activity. The pollution caused by pesticides is a significant contemporary

concern due to its adverse impact on the ecosystem. The utilization of bio-pesticides is becoming recognized as a significant approach to mitigate the ecological consequences associated with conventional chemical pesticides. While natural pesticides are commonly seen as being safer and less detrimental to the environment, their use can nonetheless have adverse effects on the ecosystem. When azadirachtin is administered at adequate doses, a significant reduction in the population of fungi and nitrifiers has been documented. However, higher concentrations of azadirachtin have been shown to have a pronounced biocidal effect on soil microorganisms and their associated activities. This phenomenon might perhaps be attributed to the presence of mutually antagonistic traits. Given that soil serves as the final repository for these substances, it is important to do research on the impacts of chemical and biopesticides on the variety of soil microorganisms and enzymes.

Soil microbial communities play crucial roles in the decomposition of organic matter, nutrient recycling, regulation of nitrogen and carbon cycles, storage and release of essential nutrients, promotion of plant growth, and serving as a significant food source at the base of the ecological food webs. Microbes play a key role in cleaning up damaged habitats by degrading organic pollutants connected to soil. Pesticide impacts on the diversity of soil microbial populations and enzymatic activity are critical to soil sustainability, agricultural production, and environmental quality. Non-target organism exposure and the off-site mobility of pesticides have emerged as major problems in the effort to maintain a pollution-free environment. Pesticides may have a detrimental effect on soil fertility. The ideal insecticide would be non-permanent, biodegradable, and lethal only to the bug it was designed to kill. When used in nature, pesticides undergo a metamorphosis. Physical, chemical, and biological factors, of which bacteria are a prominent component, are responsible for these transformations. The method of transformation, which might involve oxidation, hydrolysis, reduction conjugation, etc., is catalyzed by a wide variety of enzymes. The breakdown process is heavily influenced by the pesticide's physical and chemical properties. The degradation process involves both biotic and abiotic factors, with a lot of focus on the latter. Therefore, the soil microbe population is crucial for maintaining soil health and decomposing pesticides.

## 2. LITERATURE REVIEW

Jeffries, et.al. (2018) The escalating global concern of environmental and agricultural chemical pollution poses a significant threat to sustainability and human well-being. Among the various contaminants, organophosphorus (OP) compounds represent a substantial class that can be subject to microbial degradation. Surprisingly, no prior studies have harnessed system-wide ecogenomic tools or metagenomics to comprehensively comprehend degradation mechanisms in their natural context and predict degradation potential. Consequently, the functional genes and genetic capabilities responsible for degradation and the responses of microbial communities to contamination have remained a mystery. To bridge this critical knowledge gap, we undertook a comprehensive approach. We employed shotgun sequencing to analyze the DNA of microbial communities thriving in pesticide-treated agricultural soils. Our examination zeroed in on assessing changes in the wealth of utilitarian qualities and the ordered piece of the microbial populaces. The outcomes of our study unveiled the existence of two distinct soil groupings, each characterized by unique functional and taxonomic features. Degradation experiments showed that these groupings matched soils' organophosphorus degradation capability, with the fastest degrading community having increased transport and nutrient cycling routes and phosphorus metabolism enzymes. This occurred amid taxonomic community alterations that may have been caused by pollution adaption and exposure. Our findings illustrate the effectiveness of comprehensive, system-wide metagenomic approaches in predicting microbial degradation within ecosystems affected by pollution.

Ali, et.al. (2021) To fulfill rising food and fiber demand, agroecosystems rely heavily on pesticides. To boost productivity, tons of synthetic pesticides are used. Because 98% of pesticides impact non-target creatures, this scenario is dangerous. Studies indicate 80% of sprayed pesticides pollute the environment. This chapter examined pesticides' impact on the ecosystem, biodiversity, pollinators, food chains, and health. It was shown that pesticides degrade soil and water. Pesticides contaminate 90% of agro-land water sources and reduce soil respiration by 35% due to microbial life danger. Bioaccumulation and biomagnification of pesticides threaten aquatic and terrestrial food systems.

Pesticide exposures threaten biodiversity and species, according to studies. In Europe, insect biomass has dropped 70% and farmland birds 50% in recent decades. Europe, Australia, and North America likewise saw 42% species richness losses. Pesticide residues harm bees, reducing their ecological function. The UN warns that 40% of pollinators, including bees and butterflies, might be exterminated. Due to pesticide overuse, American and European honey bee populations have declined by 30%. Human life is harmed by food chain pollution. Ingesting these substances has caused cancer, endocrine, neurological, reproductive, and other illnesses. Many people have died from pesticide exposure. Pesticide exposure increases cancer risk and mental health issues by 25–30%, according to study. Pesticide exposure by fathers increases children's risk of leukemia, lymphoma, and brain cancer by 50%. We need to discover solutions that safeguard the environment and human health. Integrated pest control may be the only way to reduce pesticide consumption.

Wang & Cernava, (2020) Recent research have demonstrated that agrochemicals may significantly impact microbial ecosystems, especially those linked with cultivated plants. Common agricultural techniques like seed coating with fungicide-based matrices can harm up to 50% of naturally existing microorganisms. The off-target impacts of popular agrochemicals on microbial populations are still poorly understood. Agrochemical inputs are rising due to agricultural intensification and global pathogen pressure. This article briefly reviews pesticide interaction with microbial communities and discusses detrimental effects on the plant holobiont and beyond local system limits. Cumulative pesticide inputs that change microbial behavior may have unexpected impacts on geochemical cycles and should be prioritized in study. A complete study of such consequences will help us objectively choose the best food production methods for a growing global population and worsening climate. We propose three possible pesticide application methods that might be more sustainable and less harmful.

Dhananjayan, et.al. (2020) Agrochemicals are now essential for agricultural cultivation to meet rising demand. Agrochemical production, marketing, and use have expanded severalfold since the green revolution. The negative effects of agrochemicals on the environment and human health were discovered in the late 19th century. After discovering the negative effects of pesticide usage,

environmental and human toxicity were assessed. In this preview, this chapter discusses agrochemical production and usage worldwide. The bioaccumulation and biomagnification of pesticide residues throughout multiple trophic levels of the agroecosystem affected air, water, and soil. Also examined were lingering impacts on earthworms, fishes, birds, and people. Due to poor and indiscriminate agrochemical usage, especially pesticides, humans, the apex predators of agroecosystems, are severely harmed. The health effects of pesticide exposure from the environment, work, and unanticipated mishaps were discussed. The acute, sub-acute, and chronic harmful effects of pesticides were also examined from global research. Bio-monitoring studies are needed to determine the environmental dangers of agrochemicals. To acquire a more profound comprehension of the unfriendly effects of agrochemicals on both the climate and human wellbeing, we utilized bio-observing investigations, risk evaluation and the board procedures, as well as ecotoxicological data sets.

### **3. RESEARCH METHODOLOGY**

#### **3.1. Place And Duration of Study**

The review was led at two explicit areas: the Branch of Ecological Administration and Toxicology in Effurun and the Foundation of Rural Exploration and Preparing in India.

#### **Soil Sampling:**

The Institute of Agricultural Research and Training in India is home to agricultural ecosystems subjected to pesticides, where soil samples were taken for this study. There was prior use of pesticides in these habitats.

#### **3.2. Sample Selection:**

Soil samples were collected from the uppermost 0-15 cm layer of fields where pesticides had been regularly applied. These fields represented distinct agricultural plots subjected to various types of pesticides throughout time.

#### **3.3. Soil Characteristics:**



The collected soil was found to have a mostly sandy loam texture; however, it was varied. The pH of the soil fluctuated; some areas had slightly acidic soil, while others had slightly alkaline soil. The concentrations of nitrogen (N) and potassium (K) ranged from low to high, while the organic carbon content varied from low to medium. Phosphorus (P) levels were generally on the lower side.

#### **3.4. Sample Preparation:**

To guarantee consistency, the dirt examples were entirely blended and, in this way, gone through a 2 mm channel to eliminate any flotsam and jetsam or enormous particles. Microcosms were constructed using 3 kg of soil samples, which were put in sterile 5-liter containers.

#### **3.5. Moisture Adjustment:**

To guarantee uniformity, the soil moisture content in each container was adjusted using sterile deionized water to field capacity. This contributed to the replication of normal soil moisture conditions in pesticide-exposed agricultural landscapes.

#### **3.6. Incubation Period:**

Before commencing the experiment, the soil microcosms were placed in a dark environment for one week. This gave the soil time to adjust to the laboratory conditions.

#### **3.7. Pesticide Application:**

Pesticide treatments were applied to soil samples in microcosms to imitate the exposure circumstances of agricultural ecosystems. Pesticides including paraquat and carbofuran were used at the recommended rates. Paraquat, for example, was sprayed at a rate of 5 ml/kg of soil, and carbofuran at a rate of 0.1 g/kg of soil. These treatments were given weekly for eight weeks in a row, simulating regular pesticide applications in the field. The control group, on the other hand, only got sterile water.

#### **3.8. Incubation Conditions:**

Every container, including the treatment and control groups, was kept at a constant temperature of  $25 \pm 2^{\circ}\text{C}$  in a regulated setting.

### **3.9. Sampling Time Points:**

Following the eighth week of pesticide treatment, soil samples were taken for examination on a weekly or more frequent basis. These samples made it possible to evaluate changes in soil microbial diversity over time in response to pesticide exposure.

### **3.10. Determination of Soil Organic Carbon (OC):**

The study used a modified Walkley-Black technique with a colorimetric methodology to determine the amount of soil organic carbon (OC) in soil samples from pesticide-treated and control groups. A 1-gram soil test was taken from every treatment bunch, and in this way, every jar was loaded up with 10.0 ml of 1N potassium dichromate. In the wake of shaking, 20 ml of concentrated sulfuric corrosive was added to the combination. The combination was allowed to stand for 30 minutes. After digestion, each flask was supplemented with 100 ml of distilled water and 10 ml of concentrated phosphoric acid, in readiness for colorimetric analysis. Each flask was filled with three to four drops of an appropriate color indicator (such as diphenylamine) in order to measure the amount of organic carbon present. Thusly, the jars were titrated utilizing a normalized 0.5N ferrous ammonium sulfate arrangement until an adjustment of variety happened, changing from green to blue lastly to red. This change meant the total oxidation of natural carbon.

### **3.11. Analysis of Soil Enzymes**

This study aimed to investigate the impact of pesticides on microbial populations by assessing soil enzymes, including dehydrogenase, amylase, invertase, urease, protease, and phosphatase. Soil tests were gathered from both the pesticide-treated and control gatherings. Enzymatic analyses were carried out using a Hitachi (220) spectrophotometer, following established protocols for soil enzyme analysis. Dehydrogenase activity was measured to quantify soil microbial metabolic activity and changes in redox potential. Amylase, invertase, urease, protease, and phosphatase



exercises were analyzed to investigate the microbial breakdown of natural mixtures inside the dirt. Each of these enzyme activities plays a specific role in nutrient cycling and the decomposition of organic matter within the soil ecosystem. The spectrophotometer was utilized to quantify enzymatic activities by colorimetric measurements, enabling for reliable estimation of enzyme activity levels.

### **3.12. Determination of Amylase Activity:**

This study assessed amylase activity in soil samples from pesticide-exposed and control groups to understand the impact of pesticides on soil microbial enzyme activity over time. The process involved sample preparation, enzymatic assay, substrate addition, termination, and analysis. The amylase enzyme facilitated the breakdown of starch into simpler sugar compounds, and the resulting clear supernatant was subjected to analysis at 600 nm using a spectrophotometer. Control samples were prepared to assess the degree of starch hydrolysis in the absence of amylase activity, providing a baseline for comparison.

#### **Calculation:**

The research evaluated the temporal effect of pesticides on amylase activity through the use of the following formula: amylase activity ( $\mu\text{g}$  glucose released/gram soil/hour at  $37^\circ\text{C}$ ) =  $(C - C_{\text{blank}}) \times V / (\text{sw} \times \text{dwt} \times t)$ .

### **3.13. Estimation of Invertase Activity:**

This research examined soil samples to assess invertase activity and gain insights into how pesticides affect soil microbial enzyme function over time. The Ross technique was utilized to gauge invertase action, which involved the expansion of 0.1 M acetic acid derivation cradle and a 10% sucrose answer for soil treated with toluene. The mixture was then incubated at  $37^\circ\text{C}$  for a 24-hour duration, with the process being halted by the addition of distilled water. The invertase activity was quantified using the same steps as for amylase activity, with the clear supernatant collected, filtered, and analyzed at 600 nm using a spectrophotometer. Both soil amylase and invertase exercises were evaluated in microgram glucose counterparts per gram of dry soil, offering a normalized proportion of these catalyst exercises.

### ➤ **Protease**

The study assessed protease activity in soil samples to understand the impact of pesticides on soil microbial enzyme function over time. The method involved preparing 0.5 grams of soil samples from both pesticide-exposed and control groups, and using an enzyme assay. The dirt example was joined with a phosphate cushion, tyrosine, and water, trailed by hatching at 30°C for 60 minutes. The response was ended by the expansion of NaOH, acidic corrosive, and the ninhydrin reagent. The blend was in this way bubbled for 15 minutes, permitted to cool, and its absorbance was estimated at 570 nm utilizing a spectrophotometer. The protease action not set in stone by contrasting it with a standard tyrosyl leucine arrangement.

### ➤ **Urease**

The study assessed urease activity in soil samples to investigate the impact of pesticides on soil microbial enzyme function over time. The method involved adding toluene to fresh soil, adding buffer solution and urea solution, and incubating for 3 hours. The resulting mixture was filtered and measured for ammonia release using the indophenol blue method. The optical thickness was surveyed at 630 nm, and the amount of  $\text{NH}_4^+$ -N delivered was resolved utilizing a reference-aligned bend. The discoveries were accounted for as  $\text{NH}_4^+$ -N in milligrams per gram of dry soil each three-hour stretch.

### ➤ **Phosphatase**

The research sought to grasp how pesticides affect soil microbial enzyme activity over time, with a specific focus on assessing phosphatase activity in soil samples. The method involved assessing phosphatase activity using air-dried soil from both pesticide-exposed and control groups. Toluene treatment was applied, and this was trailed by the presentation of 0.1 M Tris-HCl cradle alongside 0.013 M disodium phenylphosphate in Tris-HCl cushion. The flask was incubated at 37°C for 3 hours, then centrifuged to separate the clear supernatant. The phenol was estimated by mixing the

clear supernatant with Folin's phenol reagent and a 20% (w/v) Na<sub>2</sub>CO<sub>3</sub> solution. The optical thickness was surveyed at 650 nm using a spectrophotometer. Phosphatase action was evaluated and announced as micrograms of phenol freed per gram of soil ( $\mu\text{g}$  phenol/gram of soil).

### ➤ **Dehydrogenase**

The study assessed dehydrogenase activity in soil samples to understand the impact of pesticides on soil microbial enzyme function over time. The strategy included the readiness of 1 gram of new soil from both the pesticide-uncovered and control gatherings, using the 2-3-5-Triphenyl tetrazolium chloride (TTC) decrease procedure. The dirt example was brooded at 30°C for 24 hours, trailed by extraction with concentrated methanol. The subsequent filtrate was acclimated to 50 ml, and the optical thickness was estimated at 485 nm. Dehydrogenase action was evaluated as far as the formazan focus, which was determined utilizing a standard bend of triphenyl formazan in methanol. Dehydrogenase action per gram of dry soil was communicated as milligrams of formazan per gram of dry soil each hour.

### **3.14. Evaluating the Microbial Population**

The exploration researched the impact of pesticides on microbial populaces, including microorganisms, actinomycetes, growths, nitrifying microscopic organisms, and phosphorus solubilizing microorganisms (PSM). Bacterial populations were determined through the plate count agar (PCA) technique, whereas fungal populations were assessed using Rose Bengal chloramphenicol agar. Actinomycetes populations were determined using starch casein agar, and PSM populations were assessed using Pikovskaya's medium. Nitrifying bacteria populations were determined using Ashby medium, with counts taken after seven days of incubation. The methodology employed was serial dilution plate count, with counts taken after a 48-hour incubation period.

### **3.15. Statistics and Statistical Analysis**

The information went through a two-way examination of change (ANOVA) to assess the meaning of treatment impacts, and the mean qualities were looked at using the most un-massive contrast (LSD) test.

## 4. RESULTS AND DISCUSSION

### 4.1. Analysis of Soil Physicochemical Properties

Table 1 underneath presents the consequences of the physicochemical investigation directed on the uncontaminated soil test:

**Table 1: Physical And Chemical Characteristics of The Soil Sample**

Parameter	Values
Total organic carbon (%)	7.4
pH	8.35
Nitrates (mg/kg)	26.37
Phosphates (mg/kg)	2.84
Sodium (meq/100 g)	10.52
Potassium (meq/100g)	4.87
Calcium (meq/100 g)	6.25
Magnesium (meq/100 g)	0.01
Soil particle size distribution (%)	
Loam	2.55
Sand	95.135
Clay	0.154

### 4.2. Soil Organic Carbon

Figure 1 shows significant differences in pesticide-treated soil organic carbon (OC). In the soil treated with carbofuran, the organic carbon content initially increased until day 21, after which it declined by day 28. Conversely, in the soil treated with paraquat, the organic carbon content rose from 1.62% at day 7 to 2.45% at day 14, decreased to 1.94% at day 21, and then increased again to 2.48% by day 28. Baboo et al. (30) found similar organic carbon trends. The unpolluted soil control group saw a progressive rise in total organic carbon from day 7 to day 28. The variations in organic carbon levels between pesticide treatments were statistically significant at  $P=0.05$ .

Certain herbicides break down more rapidly in soils with higher organic carbon content, possibly as a result of heightened microbial activity. The presence of organic carbon in the soil plays a role in shaping the fate of pesticides, aiding in their removal from the soil matrix.

### **4.3.Enzyme Activities**

The decision to break down protease and urease catalysts depended on their vital capabilities inside the nitrogen cycle. Also, amylase and invertase were chosen because of their significance in the dirt carbon cycle. Dehydrogenase was incorporated to check the generally speaking microbial action, and the examination of phosphatase was led to survey its importance in the phosphorus cycle. The following are the aftereffects of the catalyst examinations for soil treated with pesticides:

#### **4.4.Amylase activity**

In the carbofuran-treated soil, amylase action showed an increment from day 7 to day 14, trailed by a decline from day 21 to day 28. Paraquat-treated soil showed a persistent increment from day 7 to day 28. These variations were statistically significant, suggesting that different pesticides induced changes in starch-degrading enzymes. This phenomenon might be attributed to nutrient stress and the integration of pesticide intermediates or byproducts into microbial biomass. Certain microbial gatherings, particularly pesticide-corrupting microscopic organisms and parasites, could start the breakdown of pesticides when they are brought into the climate. Previous research has shown that certain pesticide concentrations can negatively affect amylase activity.

#### **4.5.Invertase Activity:**

In a study, invertase activity in paraquat-treated soil decreased from the 7th to the 28th day, while in carbofuran-treated soil showed variations. Inside the benchmark group, invertase action showed an underlying increment from day 7 to day 14, trailed by a downfall on day 21, and one more upswing at day 28. The variation in invertase activity was statistically significant, indicating that toxic pesticide compounds disrupted cellular structures and reduced nutrient mobilization and glucose concentration. The noticed diminishing in carbofuran-treated soil could be credited to the problematic impacts of poisonous pesticide compounds.

Fig. 3 presents the consequences of invertase movement noticed all through the review. On account of paraquat, invertase movement reliably diminished from the seventh day (35.09  $\mu\text{g}$  sucrose/g soil) to the 28th day (31.87  $\mu\text{g}$  sucrose/g soil). Conversely, there were vacillations in invertase action inside the carbofuran-treated soil, going from 26.82  $\mu\text{g}$  sucrose/g soil to 28.52  $\mu\text{g}$  sucrose/g soil. In the benchmark group, invertase action expanded from day 7 (36.75  $\mu\text{g}$  sucrose/g soil) today 14 (37.68  $\mu\text{g}$  sucrose/g soil), diminished at day 21 (31.05  $\mu\text{g}$  sucrose/g soil), and afterward expanded again at day 28 (33.83  $\mu\text{g}$  sucrose/g soil). The variety in invertase action concerning various pesticides and time focuses was genuinely huge at an importance level of  $P=0.05$ . Studies have demonstrated that invertase is a surprisingly steady compound, enduring in relationship with different soil parts. By and by, during this examination, varieties in invertase catalyst movement were seen in the carbofuran-treated soil. The decrease in movement might be credited to cell harm brought about by the poisonous mixtures present in the pesticides, as well as a decrease in supplement preparation and glucose fixation.

**Table 2: Herbicides' Effects on Organic Carbon on Various Days Following Treatment**

Day	Carbofuran-Treated Soil	Paraquat-Treated Soil	Unpolluted Soil (Control)
7	1.62%	1.68%	1.61%
14	2.53%	2.45%	2.53%
21	1.92%	1.94%	1.85%
28	2.48%	2.56%	2.40%



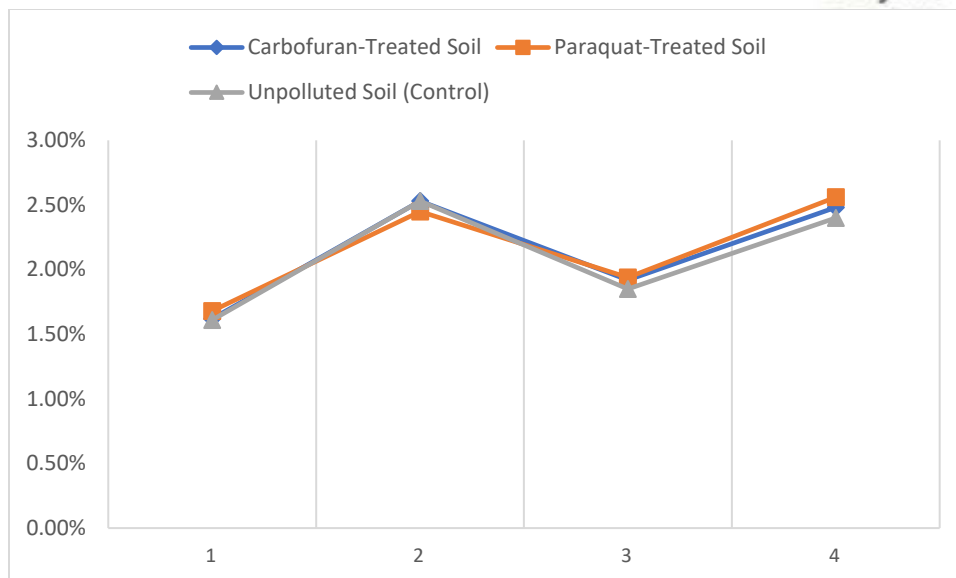


Fig. 1. Herbicide Spraying on Organic Carbon

**Table 3: Pesticides' Effects on Amylase on Various Days Following Treatment**

Enzyme	Carbofuran-Treated Soil	Paraquat-Treated Soil	Unpolluted Soil (Control)
Amylase Activity ( $\mu\text{g}$ glucose/g soil)	Day 7: 22.50	Day 7: 22.50	Day 7: 22.50
	Day 14: 23.01	Day 14: 33.32	Day 14: 23.01
	Day 21: 22.10	Day 21: 22.10	Day 21: 22.10
	Day 28: 21.21	Day 28: 33.32	Day 28: 21.21
Invertase Activity ( $\mu\text{g}$ sucrose/g soil)	Day 7: 26.82	Day 7: 35.09	Day 7: 36.75
	Day 14: 28.52	Day 14: 33.32	Day 14: 37.68
	Day 21: 26.82	Day 21: 31.05	Day 21: 31.05
	Day 28: 28.52	Day 28: 31.87	Day 28: 33.83

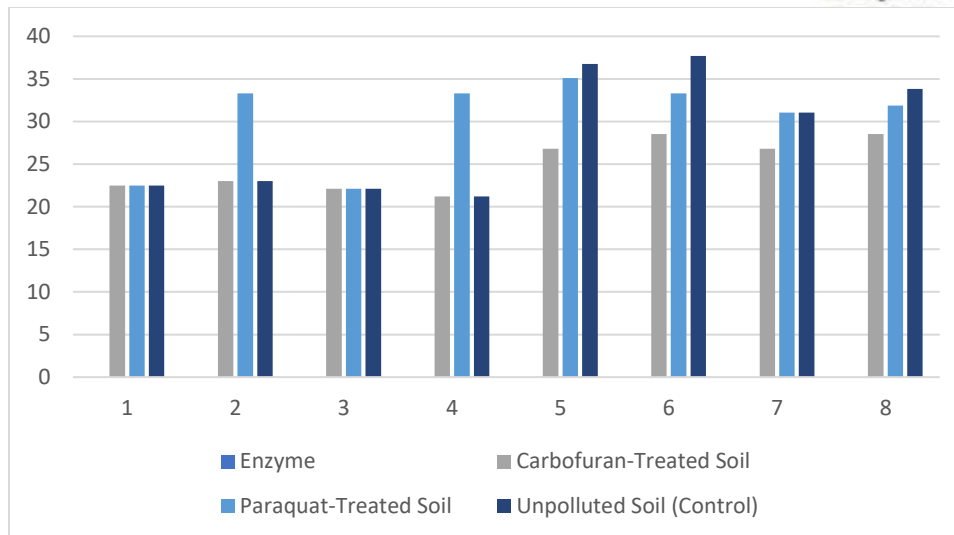


Fig. 2. Pesticide effects on amylase on different days since treatment

Table 2 compares the effects of the pesticides Carbofuran and Paraquat on amylase and invertase activities in the soil at different time points following treatment, in contrast to uncontaminated soil (Control). The findings indicate that soil treated with carbofuran keeps its amylase activity reasonably constant, with a little drop from Day 7 to Day 28, indicating no discernible effect on the activity of the enzyme that breaks down starch. The amylase activity of soil treated with paraquat increases noticeably between Days 7 and 14, then decreases at Day 21 and then increases again on Day 28. This variance might be a sign of differences in how microbes react to paraquat, which could be caused by nutritional stress or the residues of pesticides. Over a four-week period, the amylase activity of unpolluted soil (Control) remains essentially steady with just modest fluctuations, indicating no major changes in amylase activity over time. Over the course of the investigation, invertase activity in soil treated with carbofuran fluctuates without following a recognizable pattern. The impact of hazardous pesticide chemicals on cellular architecture and nutrition mobilization may be the cause of this heterogeneity. From Day 7 to Day 28, invertase activity in soil treated with paraquat consistently decreased. This suggests that paraquat may damage cellular structures, lower glucose concentration and nutrient mobilization, and ultimately cause a drop in invertase activity. In summary, the table highlights variations in the influence of carbofuran and paraquat on the activities of amylase and invertase in the soil. The significance of

examining certain enzyme activity as markers of microbial reactions to pesticide exposure in agricultural settings is highlighted by these results.

**Table 4: Pesticides' Effects on Invertase on Various Days Following Treatment**

Day	Paraquat-Treated Soil	Carbofuran-Treated Soil	Unpolluted Soil (Control)
7	35.09	26.82	36.75
14	33.32	28.52	37.68
21	46.27	26.36	37.23
28	31.87	28.52	33.83

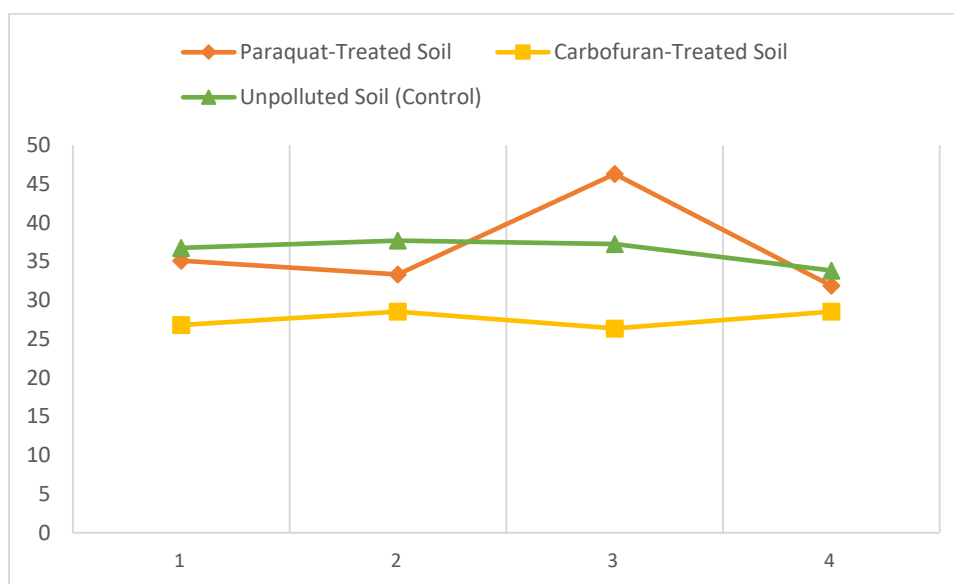


Fig. 3. Pesticide effects on invertase following treatment on various days

Table 3 shows the effects of paraquat and carbofuran on soil invertase activity during a four-week period. By Day 28, invertase activity had dropped from 35.09  $\mu\text{g}$  sucrose per gram of soil after paraquat treatment to 31.87  $\mu\text{g}$ . Paraquat may have an impact on the microbial population that breaks down sucrose by reducing invertase activity. The activity of invertase in soil treated with carbofuran varies. When compared to soil treated with paraquat, invertase activity is lower on Day 7 but rises to 28.52  $\mu\text{g}$  by Day 14. Given that invertase activity remains relatively constant at 28.52

µg by Day 28, it is possible that carbofuran has less of an impact on it than paraquat. Throughout the experiment, invertase activity in uncontaminated soil (Control) varies very little, showing that natural soil conditions free from pesticide exposure often maintain a stable level. The control group showed little fluctuations in invertase activity, indicating that pesticide exposure is the cause. In conclusion, although carbofuran varies and tends to rise, paraquat continuously lowers soil invertase activity. These results emphasize the importance of looking at enzyme activity as indicators of changes in the nutrient cycle and soil microbial population brought on by pesticides.

## 5. CONCLUSION

The motivation for undertaking this study emerges from the shortage of extensive writing on the thorough evaluation of soil quality through protein movement while pesticides are applied. The current assemblage of examination that investigates the impacts of pesticide application on soil compound action needs extensiveness, especially as far as the assortment of soil chemicals researched. Furthermore, research has confirmed that pesticides, even when administered at the rates authorized for field application, result in a significant portion infiltrating the soil. Such penetration brings about changes in the microbial populace and its circulation, alongside shifts in the physicochemical attributes of the environment. This exploration has really shown both the inhibitory and non-inhibitory effects of the pesticides under a magnifying glass on a few soil chemicals and the assessed microbial populaces. In this manner, the estimation of soil compound movement and microbial populace fills in as essential pointers in research relating to soil defilement. Hence, it is important to undertake further research endeavors that investigate additional soil characteristics, such as soil micro fauna and other enzymes, which have the potential to influence soil microorganisms, their activities, and overall soil health. These studies are necessary to comprehensively evaluate the environmental effect of pesticides.

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