

Exploring the Diversity of Beneficial Microorganisms and Their Effects on Plant Growth and Nutrient Uptake

Marada Srinivas

Research Scholar

Botany

Dr. Anita (Assistant Professor)

Guide Name

DECLARATION: I AS AN AUTHOR OF THIS PAPER /ARTICLE, HEREBY DECLARE THAT THE PAPER SUBMITTED BY ME FOR PUBLICATION IN THE JOURNAL IS COMPLETELY MY OWN GENUINE PAPER. IF ANY ISSUE REGARDING COPYRIGHT/PATENT/ OTHER REAL AUTHOR ARISES, THE PUBLISHER WILL NOT BE LEGALLY RESPONSIBLE. IF ANY OF SUCH MATTERS OCCUR PUBLISHER MAY REMOVE MY CONTENT FROM THE JOURNAL WEBSITE. FOR THE REASON OF CONTENT AMENDMENT/OR ANY TECHNICAL ISSUE WITH NO VISIBILITY ON WEBSITE/UPDATES, I HAVE RESUBMITTED THIS PAPER FOR THE PUBLICATION. FOR ANY PUBLICATION MATTERS OR ANY INFORMATION INTENTIONALLY HIDDEN BY ME OR OTHERWISE, I SHALL BE LEGALLY RESPONSIBLE. (COMPLETE DECLARATION OF THE AUTHOR AT THE LAST PAGE OF THIS PAPER/ARTICLE)

Abstract

While it stands to reason that aeration would be incredibly helpful for tank farming, very little is understood about the effects of aeration power, particularly when employing bubble flow, and their agronomic implications. In this review, social experiments and flow field perception were used to study the impact of aeration performance on plant growth. The outcomes demonstrated that there was no straight connection between aeration power and plant advancement. Inside a thin scope of aeration force (0.07-0.15 LL (- 1) NSmin (- 1)), expanding the aeration power advanced plant development. The plant development lists, but, did not significantly change above a certain range (0.15-1.18 LL (-1) NSmin (-1)). Raising the aeration intensity (1.18-2.35 LL (-1) NSmin (- 1)) also inhibited growth. Our findings highlight the need of preserving a suitable range of aeration intensity for the best possible plant development and oxygenation. Additionally, it emphasizes the necessity to balance energy consumption and operational costs as a result of rising aeration intensity in hydroponics production.

Keywords: bubble flow; dissolved oxygen; root morphology; aeration rate; image analysis technology; dry land agriculture

1. INTRODUCTION

Plants collaborate with an assortment of soil microbial networks in earthbound biological systems, laying out related associations that thus advance plant local area efficiency, subsurface biodiversity, and environment variety. The procurement of supplements by plants from heterogeneously conveyed microsites is only one of the numerous components for which these communications are fundamental. The root rummaging system, which alludes to the multiplication of roots in supplement rich microhabitats and microbially intervened components by means of a plant-organism cooperative relationship, is a particular physiological strategy that plants use in light of the spatially heterogeneously disseminated soil supplements. For instance, symbiotic partnerships between plants and soil microbes rely heavily on arbuscular mycorrhizal fungi (AMF). Both soil microbes and soil nutrient heterogeneity are known to affect plant performance, local efficiency and competitiveness. However, the potential impact of soil microbes on the effects of soil nutrient heterogeneity on plant development is poorly understood.

The assortment in the conveyance of the accessible soil supplements inside the dirt framework in a certain microhabitat is known as soil supplement heterogeneity. The lopsided dissemination of supplement assets might result from different variables, including geography, environment, and contrasts in the accessibility of microorganisms, the circulation and disintegration of litter in the dirt, and contrasts in the parent material during enduring cycles. The environment is described by far reaching spatial variety of soil supplements, which is fundamental for species conjunction and development of individual plants as well as populace construction and efficiency of networks. The capacity of the roots to scrounge in supplement rich microsites is one component supporting such an impact in plants. However, the magnitude of heterogeneity (i.e., fixed scale) and differences in supplement content (i.e., fixed contrast) are two major sources of soil nutrient heterogeneity that can affect crop development. It's a side. Both Fix Scale and Fix Contrast play important roles in the flushing component of a plant's response to nutrient fluctuations in the soil. Previous studies have shown that plants at different spatial scales can respond differently to the same variations in soil replenishment. Differences in colonization size and soil addition are not absolute factors when it comes to the effect of spatial heterogeneity on plant development. Through their activities, soil

microbes, regardless of their size and shape, can alter the effects of soil nutrient heterogeneity on plant performance.

1.1. Direct Uptake of Nutrients by Plants from the Soil

Potassium: Since potassium (K) is the most abundant cation in plant cells, it is considered a macronutrient for plants. Potassium serves as an osmoticum for cell growth and development, regulates cellular anion charges, activates chemicals, and regulates stomatal opening and closing in plants.

Potassium lack commonly influences plants developed in sandy soils, which can cause various side effects like reduced development and richness, sautéing of leaves, twisting of leaf tips, and yellowing of foliage (chlorosis).

For a long time, potassium take-up pathways have been the focal point of broad exploration. Early exploration proposed that plants straightforwardly assimilate potassium from the dirt utilizing both high and low liking transport instruments. At the point when potassium levels in the dirt are enough for plant development and advancement, low proclivity transport frameworks commonly work. Particle diversions in the root cell plasma membrane mediate a cycle that allows K^+ to move independently from regions of relatively high outer fixation into plant cells where the concentration of K^+ is lower. The availability of potassium does not appear to have a major impact on the declaration of these low-affinity carriers.

In contrast, when potassium is scarce, plants typically activate high propensity K^+ transport channels. Two proteins have been identified as the key transporters of potassium in Arabidopsis, while it is believed that several more are also involved in high-affinity transport. While AtHAK5, One of these carriers is a transporter protein, said to be a dynamic vehicle for transporting potassium to plant roots. AKT1 is a channel protein with increased love for K under conditions of potassium deficiency, likely interfering with the inactive vehicle system. Late examination has uncovered that plants have an assortment of transport frameworks that let them take potassium from the dirt and disseminate it all through their bodies. Despite the fact that there is still a lot to

comprehend about potassium take-up and movement in plants, it is obvious that the systems are complex and painstakingly controlled to empower the plant to retain sufficient potassium from the dirt in different circumstances.

Iron: Iron is essential for plant growth and development because it acts as a cofactor for proteins involved in a wide variety of fundamental metabolic processes, including photosynthesis and respiration. Iron is the fourth most common metal in the outside of the earth, yet it is much of the time a restricting component for plants since it regularly shapes insoluble buildings in high-impact soils with impartial to fundamental pH. On as numerous as 30% of soils around the world, iron limitation is respected to be a worry for plants. Plants with low degrees of iron habitually display entomb excusable chlorosis, in which the veins of the leaf are as yet green yet the in the middle between them are yellow (Figure 1). Since iron is seldom solvent in many soils, plants habitually should prepare it in the rhizosphere (the region of the dirt that encompasses and is affected by the roots) prior to bringing it inside the plant. The Strategy I and Strategy II reactions are two separate cycles that have developed that plants use to get iron from the dirt.



Figure1: Iron-deficiency chlorosis in soybean

2. LITERATURE REVIEW

2023 Abdelsattar Stevia rebaudiana is used as a sugar substitute in several countries. For this plant to be produced and be available on the market, the seed must germinate. Fertility of the soil is

decreased by ongoing cultivation without fertilizers. Helpful microorganisms improve the development and dynamic associations of *Stevia rebaudiana* in the phyllospheric, rhizosphere, and endosperm. Composts increment soil ripeness and harvest yield. Over time, soil ecosystems may get harmed by chemical fertilizers. Bacteria that encourage plant growth improve the fertility and quality of the soil, which may increase plant growth and output. Thus, biocompatible beneficial microbe injection promotes plant growth while minimizing the negative effects of artificial fertilizers. Plant development and stress resistance are enhanced by endophytic bacteria. Amino acids, polyamines, and hormones are produced by a number of plant growth-promoting microorganisms and may take the role of chemicals. Knowing how microorganisms and stevia interface powerfully can help make advantageous bacterial bio-details, use them all the more actually, and apply them to stevia to further develop creation and quality.

Bag, (2022) Due to its distinct flavour and aroma, tea (*Camellia sinensis*) is the most widely consumed and lucrative beverage in the world. Because it aids plants in acquiring nutrients from the soil and managing stress, microbiologists are currently studying the tea microbiomes. Tea roots are invaded by AMF and other beneficial bacteria, increasing the amino acids, protein, caffeine, and polyphenols in the leaves. Rhizosphere microbial biology aids in developing agricultural systems that yield a lot of food with little harm to the environment. The purpose of this paper is to explain how AMF and rhizospheric bacteria enhance plant development, tea quality, and pathogen defense by describing the microbes and their phylogeny from both produced and natural tea rhizosphere. This paper also discusses recent findings about microbes connected to tea. The tea microbiomes as a "natural resource" might aid in the sustainability of tea production, highlighting information gaps and the need for additional microbiomes study.

Beneficial plant-associated bacteria promote plant development and disease resistance, according to Compant (2010). The positive interactions between plants and microorganisms are being explored more and more in relation to climate change factors such greater CO₂, drought, and warmth. This enables testing of broad trends and the responses of different groups of plant-associated microbes to climate change. We review 135 studies on beneficial microorganisms and host plants in relation to climate change. The majority of research discovered that elevated CO₂

enhanced ectomycorrhizal and arbuscular fungi, whereas results for plant growth-promoting bacteria and endophytic fungi were inconsistent. Plant-associated bacteria typically benefited plants in high CO₂ environments. Depending on the research system and temperature range, increased temperature affected the good plant-associated microorganisms in a positive, neutral, or adverse way. Numerous studies revealed that fungi and bacteria that support plant growth improved drought-stressed plants. This review demonstrates that plant-associated microorganisms influence how plants react to climate change.

3. OBJECTIVES OF THE STUDY

The target of the review is to decide the impacts of various aeration rates on plant development, supplement take-up, and the grouping of dissolved oxygen in a supplement arrangement. The exploration will contrast aeration rates with a few plant development factors to all the more likely comprehend how aeration rates connect with leaf region, root length, Root surface area, dry weight, nitrogen content, nitrogen uptake and nitrogen utilization capacity. Moreover, to picture and break down the bubble flow field in aquaculture at shifted aeration rates, the review utilized molecule image velocimetry (PIV) techniques.

4. METHODOLOGY

4.1. Measurement and Production

The development analyze was directed in the Tottori College Plant Supplement Research center's indoor nursery with regular lighting (35.51469 N, 134.17038 E). The nursery ecological information (Beneficial Information Table S1) was accumulated from 8 Walk to 12 April 2021, when yields were being developed.

The MINA manure from Tottori College's Plant Supplement Research centre was used for this study. Table 1 shows the purpose and components of a standard MINA slurry layout. A specific chard cultivar (*Beta vulgaris* L. spp. *cicla* cv. *Seiyou Shirakuki*) was used in this study. Swiss chard seeds were sown in vermiculite. When the most prominent true leaves appeared, the seedlings were transplanted into plastic containers 580 mm long, 370 mm wide and 150 mm high. Thirty litres of

the diluted supplement arrangement (0.5 convergence of MINA manure arrangement, pH adapted to 5.0 with sulfuric corrosive (H₂SO₄) arrangement) were added to each compartment. Seedlings developed over several weeks. On day 8 of the first day of spring, seedlings were transplanted from plastic containers to growth containers with different ventilation. The lifestyle corner was a box with a width of 160mm and a depth of 200mm. The pH was adjusted to 5.0 with a sulfuric acid solution (H₂SO₄), then 3.4 L of standard MINA compost arrangement was poured into each growing tray. A development plate with a thickness of 30 mm is installed in the living area. Each growth plate had five growth holes and one vent hole, as shown in Figure 1. A pneumatic machine (model AP-40P, Yasunaga Vacuum Apparatus Inc., Tokyo, Japan) with circular aerators (i.e. 25 mm distance between aerators). Aeration rates of 0.25, 0.50, 1.00, 2.00, 4.00 and 8.00 L·min⁻¹ were applied using stones in each treatment tank. For each aeration rate (treatment), he sowed 5 seedlings in 4 replicates (growing plots). To reduce the variation in experimental results due to temperature, wind speed and light exposure in different parts of the seedbed, containers with different aeration rates were randomly placed in different positions, occasionally exchanged and repositioned. of supplements changed to full time.

Table 1: The make-up and strength of the MINA fertilizer standard solution

Composition	Concentration
Potassium nitrate (KNO ₃)	1.600 mM
Potassium dihydrogen phosphate (KH ₂ PO ₄)	0.400 mM
Calcium nitrate (Ca(NO ₃) ₂ ·4H ₂ O)	0.900 mM
Calcium chloride (CaCl ₂ ·2H ₂ O)	0.100 mM
Magnesium nitrate (Mg(NO ₃) ₂ ·6H ₂ O)	0.300 mM
Magnesium sulfate (MgSO ₄ ·7H ₂ O)	1.700 mM
Ferrous sulfate (FeSO ₄ ·7H ₂ O)	35.800 μM
Manganese sulfate (MnSO ₄ ·5H ₂ O)	9.000 μM
Boric acid (B(OH) ₃)	18.400 μM
Zinc sulfate (ZnSO ₄ ·7H ₂ O)	1.500 μM
Copper sulfate (CuSO ₄ ·5H ₂ O)	0.200 μM
Ammonium molybdate (NH ₄) ₆ Mo ₇ O ₂₄ ·4H ₂ O	0.004 μM

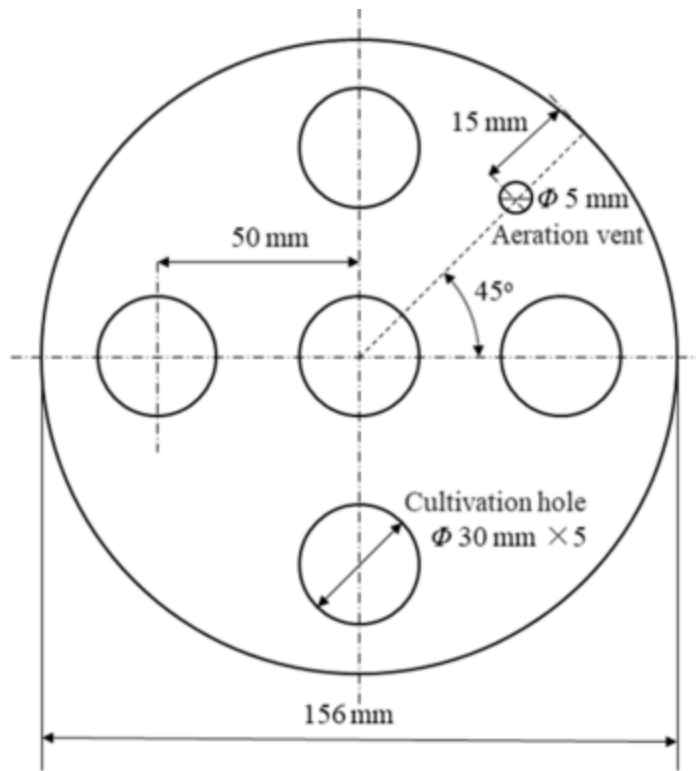


Figure 1: Size specifications for a culture plate



Figure 2: The study's hydroponic cultivation experiment

The effect of different aeration rates on the dissolved oxygen (DO) content of the auxiliary assembly in the processing chamber was investigated using a DO meter (OM-71-L1, HORIBA) to determine the dissolved oxygen content of the auxiliary assembly at various aeration rates. Shown by estimating, Tokyo Japan). Replacing the developer plate changes the amount of dissolved oxygen in the refill assembly, and opening the developer plate increases the contact area between the healthy assembly and the air. Misleading or unpredictable openings and movements, therefore, affect preliminary results, as lifestyle plates have factors other than ventilation rates. Neither the development plate was opened nor the sensor was unexpectedly inserted into the supplemental assembly during development to measure the physical and material properties of the supplemental assembly.

Plants were harvested on April 12 and separated into roots and shoots. Plants grown with varying amounts of oxygen were analyzed for leaf area, root length and surface area, dry weight, N (nitrogen) content, and N uptake using the methodology and equipment described in that study analyzed for quantity. Nitrogen use efficiency was calculated by dividing the plant's total dry load by the plant's nitrogen uptake.

5. DATA ANALYSIS

The information was examined utilizing factual analysis programming (SPSS 25, IBM, and New York, NY, USA). One-way analysis of change was used as the factual analysis strategy in this review, trailed by Duncan's numerous reach tests at $p < 0.05$. Means and standard mistake were utilized to communicate the factual discoveries.

5.1. Bubble Flow Field Visualization

A non-nosy optical laser estimation technique for flow perception is molecule image velocimetry (PIV). This study utilized a PIV gadget to definitively look at the bubble flow field in tank-farming. A similar size as the way of life compartment referenced in Segment 2.1 was made in a drab, straightforward acrylic container. Air was siphoned into the can to aerate it, and an assistant valve directed the aeration rate, which could be 0.25, 0.50, 1.00, 2.00, 4.00, or 8.00 L•min⁻¹. Selected

plants were used in this study to capture the flow field. Plants matured with different aeration rates were transferred to appropriate aeration rates to better observe the air bubble flow field. On the left side of the acrylic can was a laser (GPOL-5W, Nippon Laser) aimed at the center (see Figure 3). To record the development of several tracer particles (thickness:

1.01 g cm¹, typical molecular width: 0.55 mm) (HP20, DIAION) in bucket illuminated by laser light sheet the area of focus (return on capital invested) is shown in the bottom left corner of Figure 3a. The plant-filled bubble flow fields were photographed for ten seconds at a time. With the help of the PIV flow field computation programme (PIVlab2.40 integrated in MATLAB, Math Works), the condition-specific images were batch processed. Using an instantaneous snapshot of 600 nonstop bubble flow fields, we were able to calculate the average speed of each bubble's propagation. By comparing the mean speed appropriation guidelines of several aeration rates with the outcomes of aquaculture cultivation, we were able to understand the role that aeration rate plays in plant growth and supplement uptake. Agriculture, Public 11 times a year:

2021 NOTE Five of the 15 images and their motion were captured with a high-speed camera (FASTCAM-MAX 120KC, Photron) with a target resolution of 1024 1024 pixels and 60 frames per second. The centroid (return on investment) is shown in Figure 3a, moving clockwise from the bottom left. The flow field of the plant-filled foam was filmed for 10 seconds each. Images generated consistently for each condition were batch processed using the PIV flow field calculation software (PIVlab2.40 integrated with MATLAB, Math Works). A determination of the average velocity of each bubble flow field was obtained using high-speed mapping of 600 sustained bubble flow fields. Aqua-farming development results and mean speed dispersion guides of different aeration rates were utilized to demonstrate what aeration rate meant for plant development and supplement take-up.

Table 2: Conversion of ventilation volume to ventilation intensity

Aeration Rate (L-min-1)	0.25	0.50	1.00	2.00	4.00	8.00
Aeration Intensity (L.l-1 NS min-1)	0.07	0.15	0.28	0.58	1.18	2.36

6. RESULTS

6.1. Variation in Saturation and Dissolved Oxygen Concentration as a Function of Aeration Intensity

The effect of aeration force on the dissolved oxygen content of the auxiliary assembly was simulated by testing the dissolved oxygen content of the auxiliary assembly at various aeration capacities prior to construction.

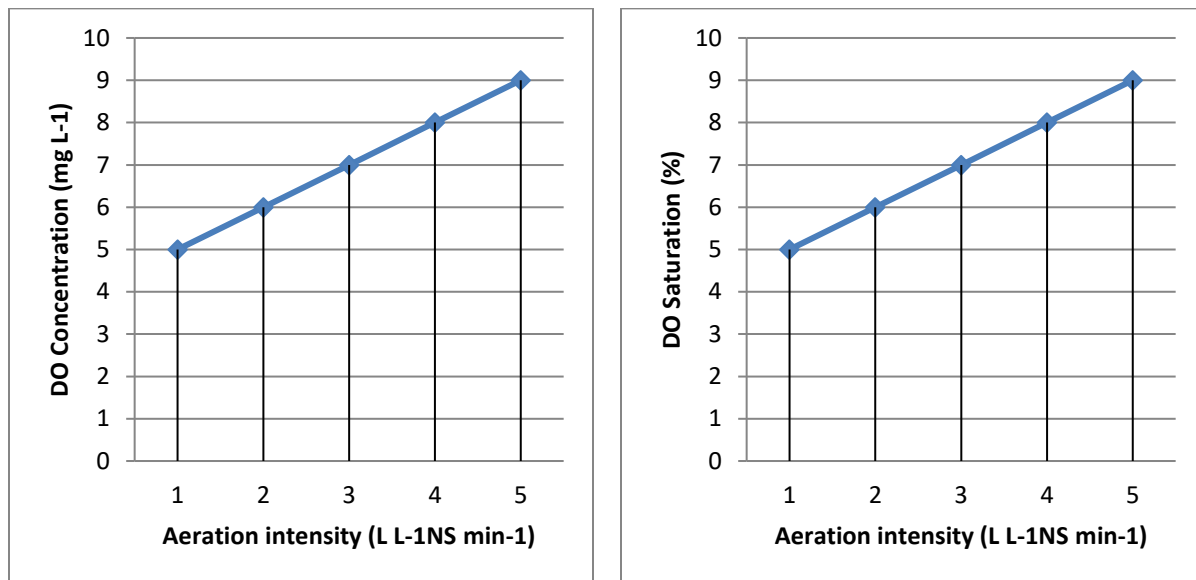
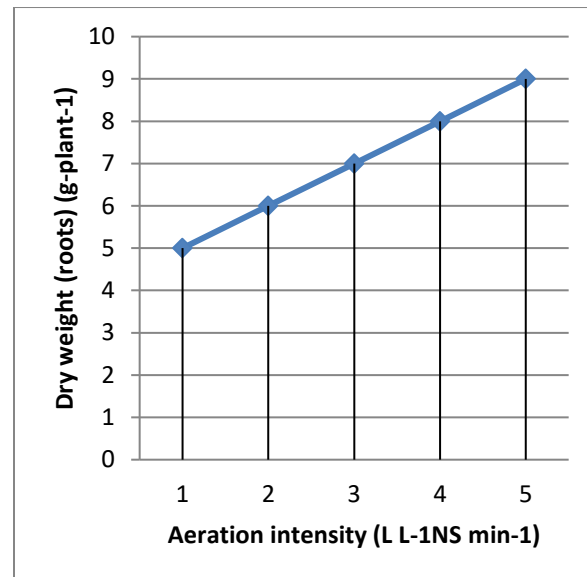
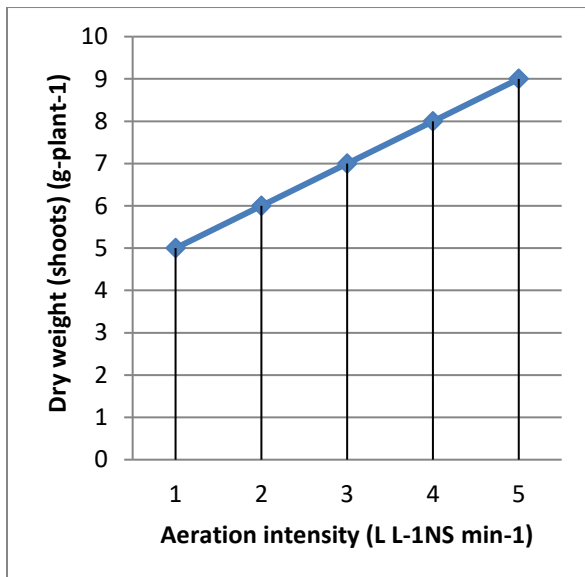


Figure 4: Prior to development, dissolved oxygen (DO) focus and immersion were estimated in this review at different aeration powers: (a) DO fixation; (b) DO immersion. Ten estimations were directed all the while (9:00-10:00 a.m. on different days; water temperature: 21°C-1°C). Information are communicated as MSE (n = 10); bars with different letter marks mean massive contrasts (p 0.05).

6.2. Effect of Root Morphology and Plant Growth on Aeration Intensity

Because they serve as primary markers of plant development, dry weight, leaf area, and water content are difficult to ignore when determining whether or not a treatment significantly influences

plant development. Plant dry weight, leaf area, and water content at different aeration levels are shown in Figure 5. These progress records show virtually identical displacement trends with increasing aeration levels. Growth data ranging from 0.07 to 0.15 L L1 NS min1 increased with aeration power. There was no significant difference in these growth records between 0.15 L L NS min and 1.18 L L NS min. These growth rates decreased with increasing aeration power ranging from 1.18 to 2.35 L·L 1 NS·min1. These development files had the lowest density at 0.07 L L 1 NS min1 and the highest density at 0.59 L L 1 NS min1.



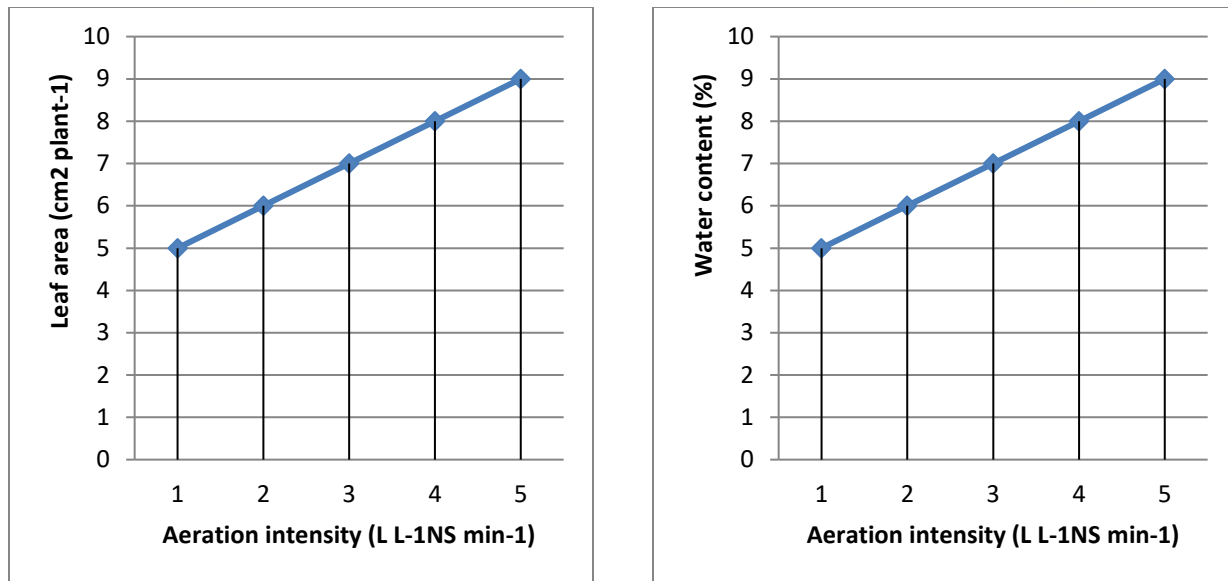


Figure 5: Under different aeration powers, the plant's turn of events and root morphology were inspected for (a) Bud dry weight, root dry weight, leaf area, water content. Data are presented as mean standard error (n=4) and highly significant differences ($p < 0.05$) are indicated by bars marked with different letters.

7. DISCUSSION

This investigation sheds light on the relationship between aeration rates in hydroponic systems and plant development and nutrient uptake. The focus of the discussion is on how low dissolved oxygen levels can result in chlorosis, stunted growth, and nutritional deficiencies in plants. It emphasizes how important it is to put technical solutions in place to ameliorate these circumstances, particularly in dry land regions where low dissolved oxygen concentrations may be more frequent. The study acknowledges the benefits of aerating nutrient solutions in hydroponics, in line with past research. Aeration techniques like blending and air gurgling are believed to be useful for oxygenating the nourishing arrangement, especially when root breath is dynamic and the water temperature is high. In the conversation, it is brought up that little is known about the effects of aeration, particularly in solutions circulated by bubble flow. It emphasizes the importance of understanding how different aeration rates affect dissolved oxygen content and solution flow rate. The discussion emphasizes the movement of the fertilizer solution as a distinguishing

characteristic of hydroponics. It alludes to past investigations that demonstrate the upsides of supplement arrangement flow on plant development and characteristics these benefits to mechanical excitement that advances root stretching and supplement take-up. According to this, aeration may also enable nutrient solutions to flow, especially through bubble flow, which promotes turbulent diffusion and provides the ideal type of physical stimulation for root growth.

The study's findings are reported in the discussion, which demonstrates that an appropriate aeration rate that falls within a specific range results in increased plant development. However, after a certain point, further increases in aeration intensity have little effect on plant growth and might even harm performance. The negative impacts of high aeration intensity's excessive flow, which hinders plant development due to excessive mechanical stimulation, are highlighted. Overall, the discussion emphasizes the need of maintaining equilibrium in hydroponic systems between the rate of aeration, the quantity of dissolved oxygen, and the flow of the solution. It emphasizes the need for additional research to determine the best aeration conditions that promote plant growth and nutrient absorption while preventing negative effects brought on by an excessive flow.

8. CONCLUSION

In this exploration, culture tests were utilized to research the impacts of aeration force on plants. Furthermore, from a design and image analysis point of view it was a curiosity made understandable by employing a liquid representation method (molecular image velocimetry). It was discovered that plant growth did not linearly increase with increasing aeration force. Expanding aeration power is more helpful for crop development when it is similarly low. How much dissolved oxygen and plant improvement, nonetheless, don't give off an impression of being advanced by expanding aeration force after a specific point. Aeration power can be changed inside a satisfactory reach to advance plant improvement. These discoveries are worthwhile for plant advancement and tank-farming development the board since they forestall the use of a pneumatic machine with an exorbitantly high power yield and furthermore stay away from the expense rise related with using an aeration force that is too high to ever be successful.

REFERENCES

1. Herman, D.J.; Firestone, M.K.; Nuccio, E.; Hodge, A. Interactions between an arbuscular mycorrhizal fungus and a soil microbial community mediating litter decomposition. *FEMS Microbiol. Ecol.* 2012, 80, 236–247.
2. Bani, A.; Pioli, S.; Ventura, M.; Panzacchi, P.; Borruso, L.; Tognetti, R.; Tonon, G.; Brusetti, L. The role of microbial community in the decomposition of leaf litter and deadwood. *Appl. Soil Ecol.* 2018, 126, 75–84.
3. Reganold, J.P.; Wachter, J.M. Organic agriculture in the twenty-first century. *Nat. Plants* 2016, 2, 15221.
4. Laliberté, E. Below-ground frontiers in trait-based plant ecology. *New Phytol.* 2017, 213, 1597–1603.
5. Jones, D.L.; Magthab, E.A.; Gleeson, D.B.; Hill, P.W.; Sánchez-Rodríguez, A.R.; Roberts, P.; Ge, T.; Murphy, D.V. Microbial competition for nitrogen and carbon is as intense in the subsoil as in the topsoil. *Soil Biol. Biochem.* 2018, 117, 72–82.
6. Capek, P.; Manzoni, S.; Kaštovský, E.; Wild, B.; Diáková, K.; Bárta, J.; Schneckler, J.; Biasi, C.; Martikainen, P.J.; Alves, R.J.E.; et al. A plant–microbe interaction framework explaining nutrient effects on primary production. *Nat. Ecol. Evol.* 2018, 2, 1588–1596.
7. Jacoby, R.; Peukert, M.; Succurro, A.; Koprivova, A.; Kopriva, S. The role of soil microorganisms in plant mineral nutrition-current knowledge and future directions. *Front. Plant Sci.* 2017, 8, 1617.
8. Jiang, F.; Zhang, L.; Zhou, J.; George, T.S.; Feng, G. Arbuscular mycorrhizal fungi enhance mineralisation of organic phosphorus by carrying bacteria along their extraradical hyphae. *New Phytol.* 2021, 230, 304–315
9. Bano, S.; Iqbal, S.M. Biological nitrogen fixation to improve plant growth and productivity. *Int. J. Agric. Innov. Res.* 2016, 4, 597–599
10. Zhang, Z.; Yuan, Y.; Liu, Q.; Yin, H. Plant nitrogen acquisition from inorganic and organic sources via root and mycelia pathways in ectomycorrhizal alpine forests. *Soil Biol. Biochem.* 2019, 136, 107517.

11. Pellitier, P.T.; Zak, D.R. Ectomycorrhizal fungi and the enzymatic liberation of nitrogen from soil organic matter: Why evolutionary history matters. *New Phytol.* 2018, 217, 68–73
12. Baiyin, B.; Tagawa, K.; Yamada, M.; Wang, X.; Yamada, S.; Yamamoto, S.; Ibaraki, Y. Effect of Substrate Flow Rate on Nutrient Uptake and Use Efficiency in Hydroponically Grown Swiss Chard (*Beta vulgaris* L. ssp. *cicla* ‘Seiyou Shirokuki’). *Agronomy* 2021, 11, 2050.
13. Baiyin, B.; Tagawa, K.; Yamada, M.; Wang, X.; Yamada, S.; Shao, Y.; An, P.; Yamamoto, S.; Ibaraki, Y. Effect of Nutrient Solution Flow Rate on Hydroponic Plant Growth and Root Morphology. *Plants* 2021, 10, 1840
14. Hussain, A.; Iqbal, K.; Aziem, S.; Mahato, P.; Negi, A.K. A review on the science of growing crops without soil (soilless culture)—A novel alternative for growing crops. *Int. J. Agric. Crop Sci.* 2014, 7, 833.
15. Wang Q, Zhang WJ, He LY, Sheng XF (2018) Increased biomass and quality and reduced heavy metal accumulation of edible tissues of vegetables in the presence of Cd-tolerant and immobilizing *Bacillus megaterium* H3. *Ecotoxicol Environ Saf* 148:269–274

Author’s Declaration

I as an author of the above research paper/article, hereby, declare that the content of this paper is prepared by me and if any person having copyright issue or patent or anything otherwise related to the content, I shall always be legally responsible for any issue. For the reason of invisibility of my research paper on the website/amendments/updates, I have resubmitted my paper for publication on the same date. If any data or information given by me is not correct, I shall always be legally responsible. With my whole responsibility legally and formally I have intimated the publisher (Publisher) that my paper has been checked by my guide (if any) or expert to make it sure that paper is technically right and there is no unaccepted plagiarism and the entire content is genuinely mine. If any issue arise related to Plagiarism/Guide Name/Educational Qualification/Designation/Address of my university/college/institution/Structure or Formatting/ Resubmission / Submission / Copyright / Patent/ Submission for any higher degree or Job/ Primary Data/Secondary Data Issues. I will be solely/entirely responsible for any legal issues. I have been informed that the most of the data from the website is invisible or shuffled or vanished from the data

base due to some technical fault or hacking and therefore the process of resubmission is there for the scholars/students who finds trouble in getting their paper on the website. At the time of resubmission of my paper I take all the legal and formal responsibilities, If I hide or do not submit the copy of my original documents (Aadhar/Driving License/Any Identity Proof and Photo) in spite of demand from the publisher then my paper may be rejected or removed from the website anytime and may not be consider for verification. I accept the fact that as the content of this paper and the resubmission legal responsibilities and reasons are only mine then the Publisher (Airo International Journal/Airo National Research Journal) is never responsible. I also declare that if publisher finds any complication or error or anything hidden or implemented otherwise, my paper maybe removed from the website or the watermark of remark/actuality may be mentioned on my paper. Even if anything is found illegal publisher may also take legal action against me

Marada Srinivas
Dr. Anita (Assistant Professor)
